

Seed-borne fungal pathogens of vegetable crops

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airborne Pathogens of Vegetable Crops

- Long distance spread of new pathogens or races of pathogens
- Low rates of infection -0.01% can result in nearly 100% infection in densely planted transplant beds or greenhouse seedling production
- Management depends on seed producer
 - Seed production field
 - Detection
 - Seed treatments

Gummy stem blight of melon



Seed-borne vs. seed transmitted organisms

- Gain access to seed
 - Active infection with in seed
 - Passive-on seed surface or contaminant
- Survive commercial processes
 - Harvest, cleaning, treatment, storage
- Establish on emerging seedlings
 - Primary or secondary transmission

active seed infection

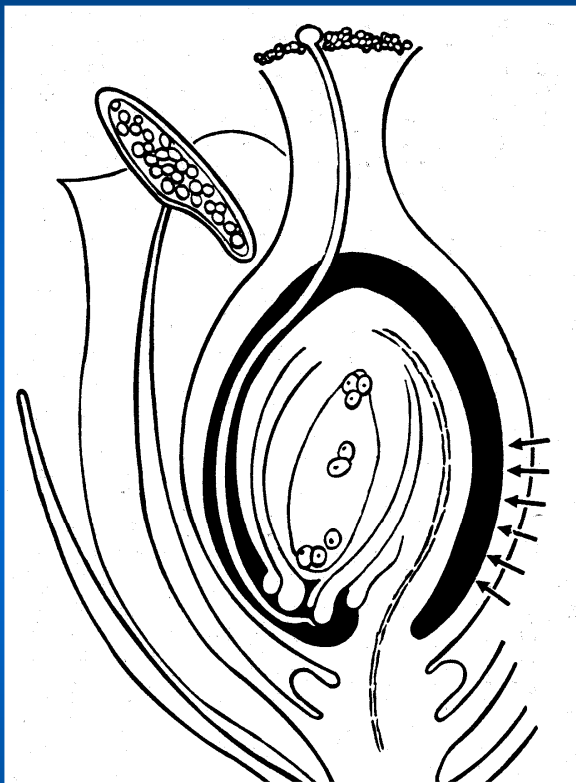
A. Penetration through ovary wall vascular system

B. Systemic infection via

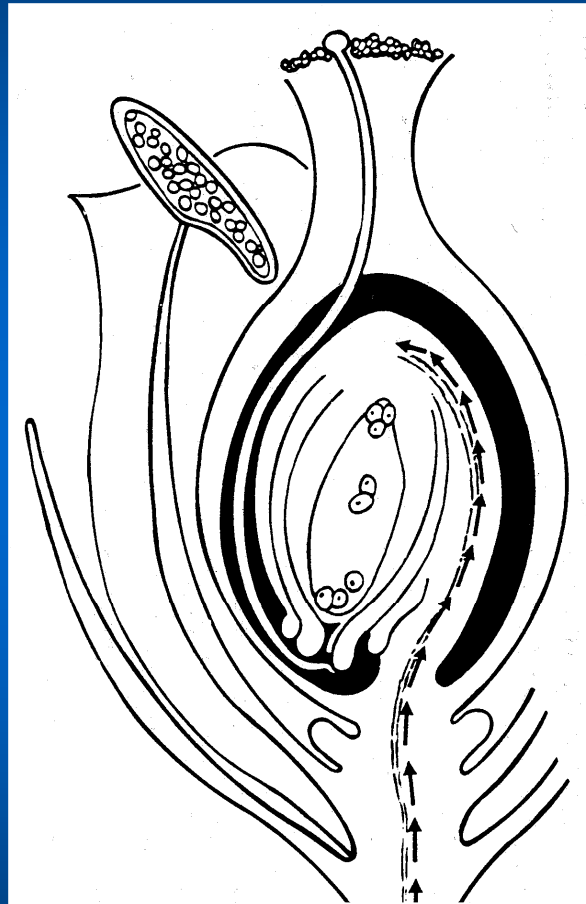
vascular system

C. Penetration through

floral parts



E.g.: *Botrytis aclada*, *B. allii* (onion)
Cladosporium variabile (spinach)



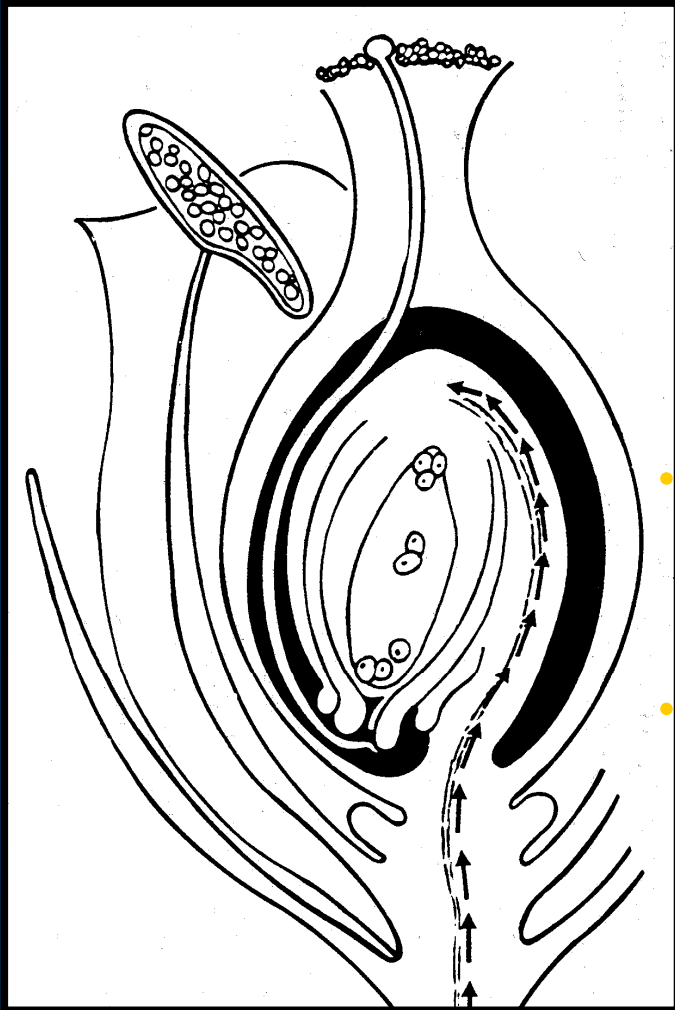
E.g.: Vascular wilt fungi, endophytes



E.g.: *Ustilago nuda* (grains)
Cucumber mosaic virus

From Maude (1996)

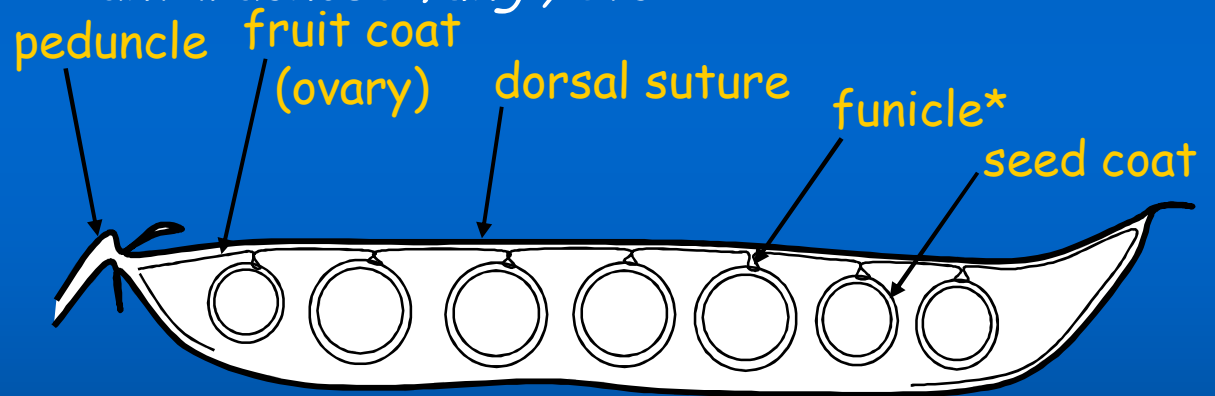
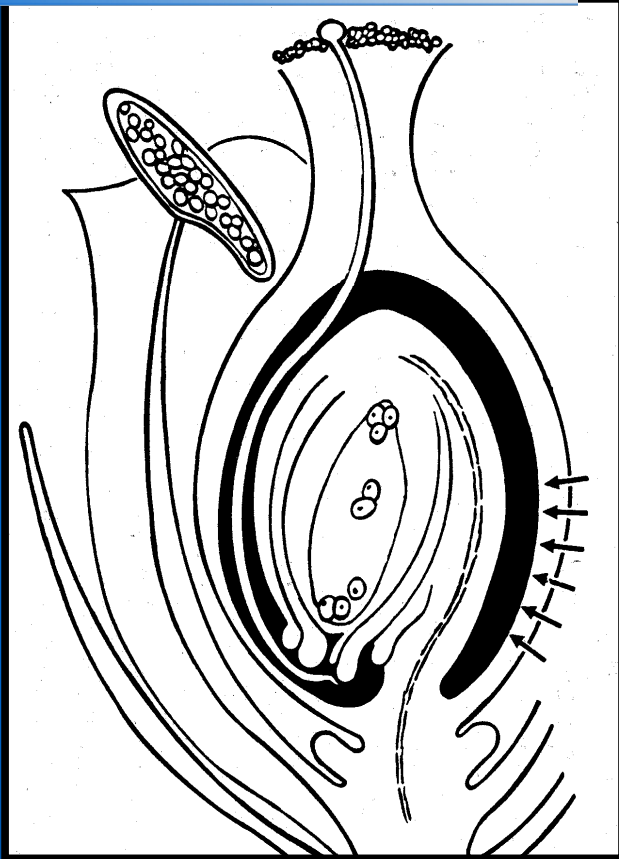
Systemic infection via vascular system



- Some fungi, e.g.:
 - Vascular wilts (*Verticillium dahliae*, *Fusarium oxysporum*)
 - some downy mildews
- Direct connection between embryonic & endospermic tissue becomes disconnected as seed develops
- Potential for transmission affected by degree of internal infection

Infection via flower or fruit

- Weak necrotrophs:
 - *Botrytis cinerea* - infected petals remain attached to developing fruit
- Aggressive necrotrophs:
 - Attack floral parts directly
 - e.g., *Ascochyta pisi*, *Alternaria brassicicola*, *Leptosphaeria maculans*, anthracnose fungi, etc



- Fleshy fruits (e.g., Solanaceae) - seed attached to central placenta
 - infect via calyx - placenta - funicle - embryo
- Umbelliferae & Liliaceae - flowers exposed in umbels
- Seed transmission is typically discontinuous (infection outside embryo)
 - affected by environmental conditions

Brassica seed



Alternaria brassicicola & *A. brassicae*

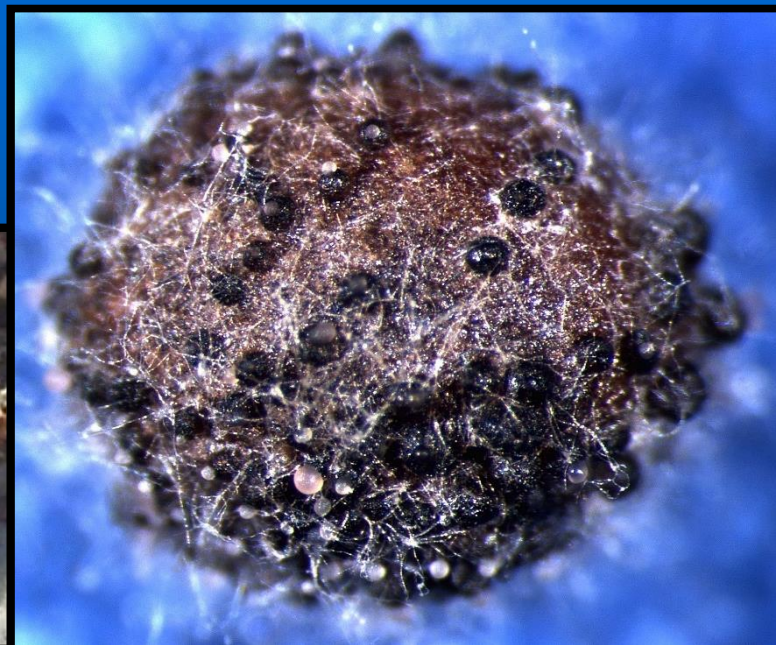
Correlation of severity of pod spot with:

Seed germ = -0.89, Seed rot = +0.88

du Toit & Derie, 2003. Fung. & Nem. Tests 58:V026



Leptosphaeria maculans



Botrytis scape & flower blight

of onion: *B. aclada* *B. allii*

B. squamosa

B. byssoidea





Seedborne *Botrytis aclada* & *B. allii* in onion



Fungal Seed-borne Pathogens

“ Alliaceae-Onion/Leek

- . Botrytis neck rot- Botrytis aclada

“ Amaranthaceae- Beet/Swiss chard

- . Phoma damping-off/canker-Phoma betae
- . Downy mildew- Peronospora farinosa f.sp. betae

“ Amaranthaceae- Spinach

- . Downy mildew- Peronospora farinosa f.sp. Spinaciae
- . Fusarium wilt- Fusarium oxysporum f.sp. Spinaceae
- . Verticillium wilt- Verticillium dahlia
- . Anthracnose- Collectotrichum dematatum
- . Cladosporium leaf spot-Cladosporium variable
- . Stemphyllium leaf spot-Stemphyllium botryosum
- . White rust- Albugo occidentalis

Major Fungal Seed-borne Pathogens

” **Apiaceae**-Carrot/Parsnip

- . Black rot/Black crown-*Alternaria radicina*
- . *Alternaria* leaf blight-*Alternaria dauci*
- . *Cercospora* leaf blight-*Cercospora carotae*
- . Similar pathogens on Celery/Parsley/Cilantro

” **Asteraceae**-Lettuce

- . *Septoria* leaf spot- *Septoria lactucae*

” **Brassicaceae**-cabbage, etc

- . Black leg- *Leptosphaeria maculans* (*Phoma lingam*)
- . Black spot- *Alternaria brassicola*
- . *Alternaria* leaf spot-*Alternaria brassicae*
- . White rust-*Albugo candida*

Major Fungal Seed-borne Pathogens

“ Cucurbitaceae-Cucumber/Squash/Melon

- . Fusarium crown and foot rot-F.solani f.sp. cucurbitaceae
- . Gummy stem blight- Didymella bryoniae

“ Solanaceae-Tomato

- . Didymella stem rot-D. lycopersici
- . Early blight-Alternaria tomatophila/solani
- . Fusarium crown and root rot-F. oxysporum f.sp. radices-lycopersici
- . Fusarium wilt-F. oxysporum f.sp. Lycopersici
- . Late blight- Phytophthora infestans
- . Phoma rot- P. destructive var destructive
- . Septoria leaf spot- S. lycopersici

Major Fungal Seed-borne Pathogens

“ Malvaceae-Okra

- . Anthracnose-Colletotrichum dematium
- . Foot and root rot-Fusarium oxysporum
- . Corynespora leaf spot-C.cassicola

“ Solanaceae-Eggplant

- . Anthracnose-Colletotrichum sp.
- . Phomopsis fruit rot- P. vexans

“ Solanaceae-Pepper/Chili

- . Anthracnose-Colletotrichum sp.

ment-Seed Producer

- “ National/International Quarantine Rules
- “ Seed Production Field selection
 - . Produce seed in low rainfall areas- avoid overhead irrigation- many pathogens are dispersed by splashing water
 - . Use plastic mulch/trellis to keep fruit off soil
 - . Appropriate crop rotation/ weed host control
 - . Avoid field with history of persistent soil pathogens such as Fusarium/Verticillium wilt fungi

Management-Seed Producer

- “ Plant pathogen free seed in seed production field
 - . indexed for pathogen (culture, visual, ELISA, PCR)
 - “ Amount of seed tested determined by allowable threshold- need better research
 - . Hot water treatment
 - . Fungicide treatment
 - . Other treatment-essential oils, biologicals



reatment for freeing seed of bacterial and fungal infection

- “ May be deleterious to %weak seed+ test small sample first
- “ May reduce seed storage life
- “ Need precise temperature control
- “ Cannot use on cucurbit/legume seeds
- “ Amaranthaceae-20-25 min @ 50-52 °C
- “ Apiaceae-20 min @ 50 °C
- “ Asteraceae-30 min @ 48°C
- “ Brassicaceae-20-25 min @ 50 °C
- “ Solanacease 25 min @ 50 °C

Other Physical Methods

“ Aerated steam

- . Forsberg et al., 2002. J. Plant Diseases and Protection 109: 357-370
- . Amein et al., 2011. J. Plant Diseases and Protection 118: 214-221

“ Microwave (electron)

- . Tinivella et al., 2009. European J. Plant Pathology 123: 139-151
- . Amein et al., 2011. J. Plant Diseases and Protection 118: 214-221

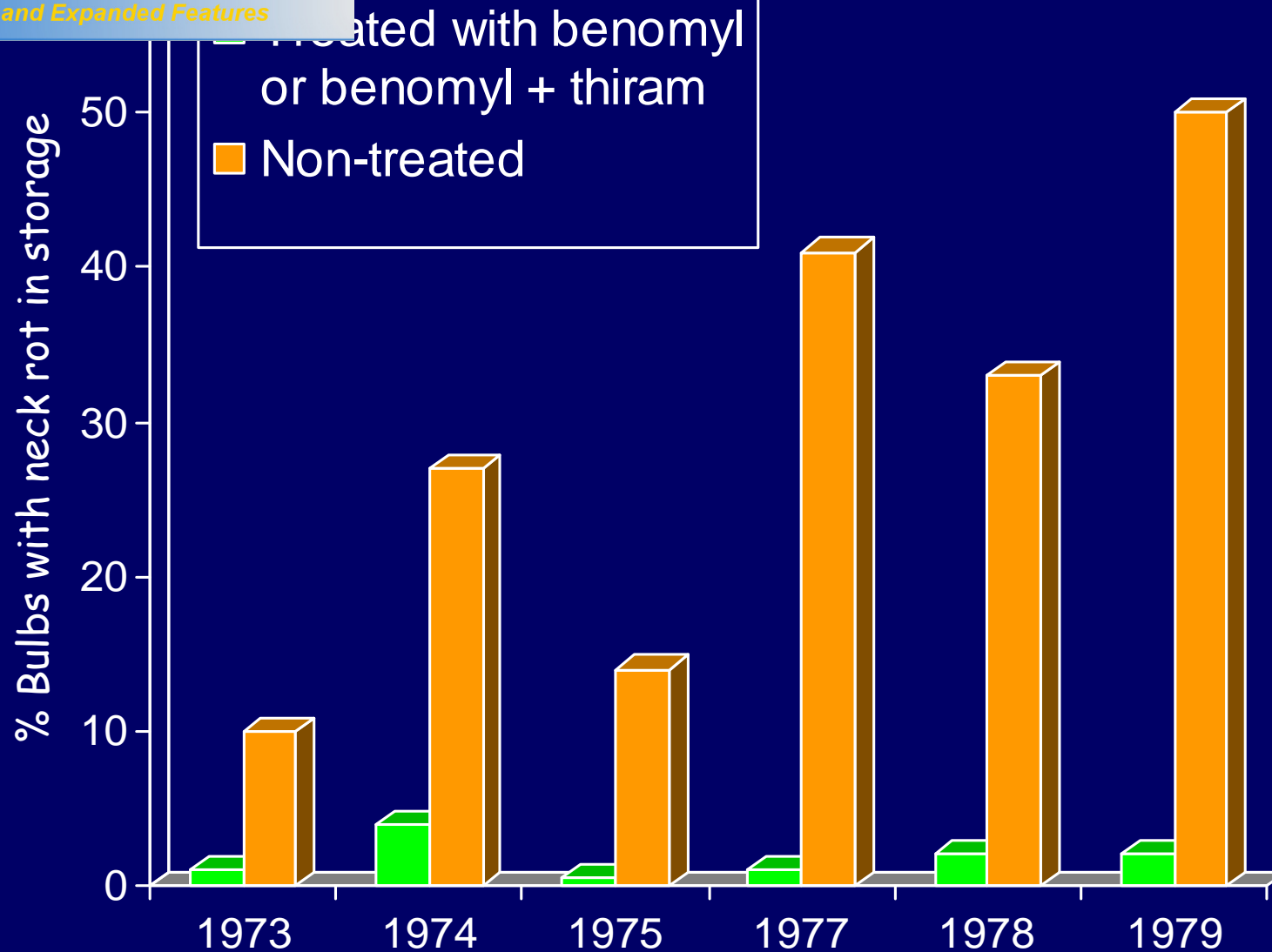
Fungicides and other treatments

“ Fungicide treatments

- . Seed soaks . Thiram/systemic fungicides (benzimidazole, triazole, SDHI, QoI (strobilurin))
- . Traditional seed treatments-systemic fungicides

“ Other

- . Use of essential oils . Thyme oil, garlic oil, etc
- . mycofumigation- *Muscodor albus*
- . Bleach, peroxyacetic acid, chlorine dioxide or other disinfectants
- . Biopesticides-*Bacillus* sp., *Pseudomonas* sp., *Trichoderma* sp., *Streptomyces* sp.- many commercial products



Control of neck rot in stored onion bulbs in the UK using seed treatments (from Maude, 1983. Seed Sci. & Technol. 11:829-834)

relative efficacy

Pathogen	Fungicide	Physical means	Plant Extracts /oils	BCA	BCA + Oils
Carrot Alternaria dauci	+++	+ / +++	++	+ / ++	?
Cabbage Alternaria	+++	+ / +++	++	++	++
Cabbage Black leg	+++	+ / +++	++	- / ++	++
Tomato Fusarium wilt	+++		++ / +++	++	++
Legume Ascochyta	+++	-	++	+	

+++ >80%, ++=50-80%, +=20-50%

Pest Management Science
70:860-868 2014.

Management-Seed Producer

- “ Use appropriate fungicide/insecticide programs on seed crop
- “ Visual inspection of seed production field for sign/symptoms of disease
- “ Timely harvest
- “ Seed cleaning- remove contaminating pathogens
- “ Seed testing for pathogen infection
 - . Int.Seed Health Initiative-Veg-2002 Seed Health Testing Methods Reference Manual
 - . ISTA-Manual for Seed Health Testing Methods
 - “ I-Validated Test Methods
 - “ II Peer Reviewed Methods

Reference websites

With new technologies methods change rapidly

“ International Seed Testing Association

. <http://www.seedtest.org/en/home.html>

“ International Seed Health Initiative

. <http://www.worldseed.org/ifs/ishi.html>

Management

Transplant/Seedling Producer

- “ Purchase only high quality seed with appropriate treatment
- “ Use Bacillus, Pseudomonas, Trichoderma seed soil treatments
- “ Avoid splashing water during irrigation-fine mists or use watering mats
- “ Do not work amongst or handle wet plants
- “ Scout regularly for signs of damping-off, leaf blights, etc-
remove and isolate
- “ Use appropriate fungicide sprays-fungicide resistance
management
 - . azoxystrobin/Gummy stem blight melon

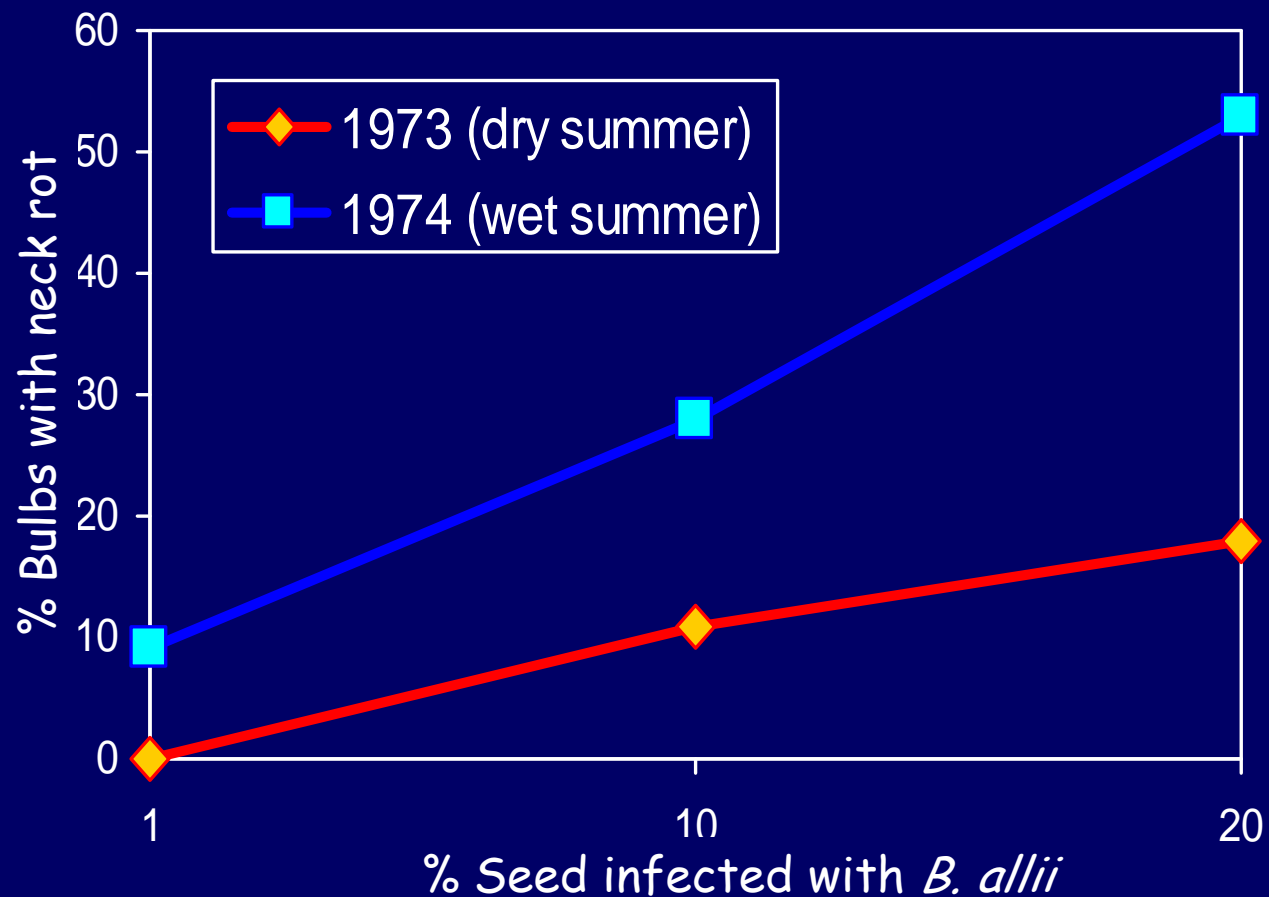
for seed transmission

- **Environmental factors- seed production field**
 - temperature*
 - moisture* - many seedborne pathogens are water splash disseminated-avoid overhead irrigation, low rainfall areas
 - degree of internal infection of seed
- **Genotype**
 - host resistance, maternal vs. paternal
 - pathogen isolate/strain
- **Inoculum potential**
 - amount of inoculum & conduciveness of environment
 - transmission rate
 - threshold(s) for seedborne inoculum
 - complex, lack of adequate field research

environment on seed transmission of *B. allii* in onion



Tichelaar, 1967; Maude & Presly, 1977a



Relationship between infected onion seed & post-harvest neck rot in the UK (from Maude & Presly, 1977b)

Ellerbrock & Lorbeer (1977): Field infection sources were more important in NY than infected seed.

Seedborne pathogens: Considerations for risk analysis

- **Epidemiology of the pathogen**
 - Potential for seed transmission
 - Conditions for disease development & spread
 - Ease of eradication
- **Alternative sources of inoculum**
 - Infested residues
 - Soilborne inoculum
 - Infected adjacent or overwintering crops or weeds
- **In-field control measures**
 - Resistant cultivars
 - Fungicides & forecasting
 - Crop rotation
 - Plant spacing, row orientation, direct seeding
 - Irrigation (system & schedule)
- **Threshold(s) for seedborne inoculum**
 - Environmental conditions?
 - Resistant vs. susceptible cultivars?

Seed pathogens: Implications

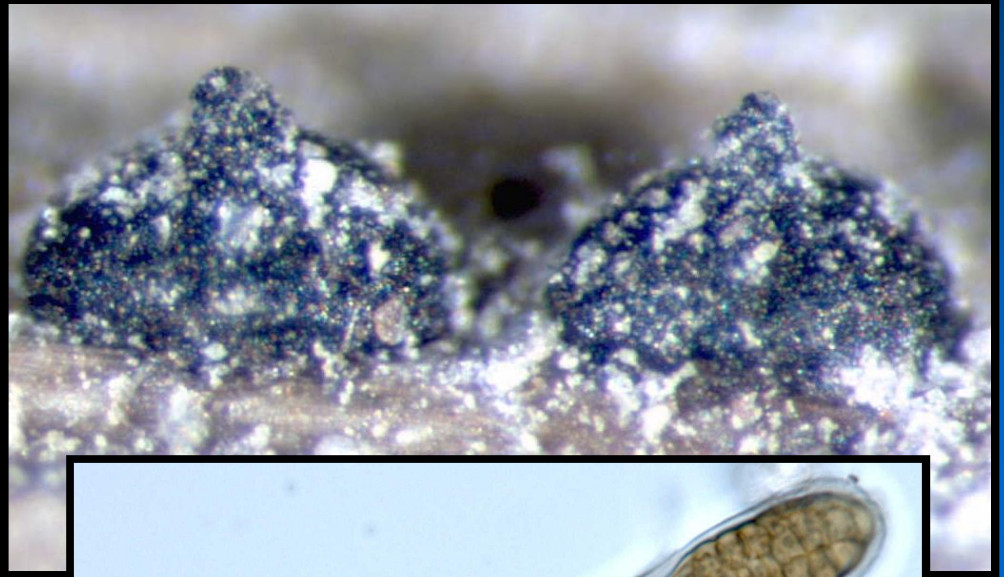
Alternative inoculum sources for seedborne carrot diseases

- Infested residues (*Alternaria dauci* in CA) (Gilbertson et al.)
- Soilborne inoculum (*Alternaria radicina* = 8 years)
- Infected adjacent or overwintering crops or related weed hosts (*X. campestris* pv. *carotae* in carrot seed crops in PNW)



du Toit et al., 2005. Plant Dis. 89:896-907.

Sources of inoculum for *Stemphylium* blight in spinach seed crops



Overwintering fruiting bodies only on residues on soil surface

du Toit & Derie, 2003. Phytopathology 93:S22

gates presented by seed borne pathogens

- Determine definitively if a seedborne pathogen is seed transmitted?
- Develop thresholds for seedborne inoculum, applicable to a range of environments?
- Regulatory implications for seed transmitted vs. seedborne status?

